BrainXell Microglia Monoculture
Protocol (v10.2)

CONTENTS

- One vial of cryopreserved human iPSC-derived Microglia (BX-0900, BX-09XX-XX)

SAFETY PRECAUTIONS

CAUTION: The use of full personal protective equipment (PPE) is strongly recommended when handling or transferring cryovials including safety-rated goggles, face shield, cryogenic gloves, and a lab coat. Any cryovial stored in liquid nitrogen has the potential to rapidly expand upon warming and carries the risk of rupture or explosion. Avoid submerging the vial cap in liquid nitrogen to reduce risk.

STORAGE

- Immediately transfer the cryovial of microglia to a liquid or vapor nitrogen storage system.

ADDITIONAL MATERIALS NEEDED

- DMEM/F-12 Medium, +L-glutamine +HEPES (Thermo Fisher Scientific #11330032)
- B-27 Supplement (Thermo Fisher Scientific #17504044)
- N-2 Supplement (Thermo Fisher Scientific #17502048)
- GlutaMAX Supplement (Thermo Fisher Scientific #35050061)
- MEM NEAA (Thermo Fisher Scientific #11140050)
- Chemically Defined Lipid Concentrate (Thermo Fisher Scientific #11905031)
- Ascorbic Acid (MilliporeSigma #A8960)
- Geltrex (Thermo Fisher Scientific #A1413201) or Cultrex (R&D Systems #3432-001-01)
- M-CSF (Thermo Fisher Scientific #300-25)
- IL-34 (Thermo Fisher Scientific #200-34)
- TGF-B1 (Thermo Fisher Scientific #100-21C)
- PDL-Coated 96-Well Plates (Refer to BrainXell PDL Plate Coating Protocol)

PROCEDURE

Day 0 Seeding Preparation

To seed (1) full 96-well plate you will need 2.2 – 3.3 million live cells. Additional cells may be used depending on individual research needs. Review the Certificate of Analysis (CoA) for viability and seeding information.

1. Gather the Microglia Base Medium components according to the recipe (see Media Compositions section below).
    a. Note that M-CSF, IL-34, and TGF-β1 are lyophilized powders. Follow the manufacturer's instructions for reconstitution and long-term storage. We recommend creating stock solutions of 100 µg/mL for M-CSF, 100 µg/mL for IL-34, and 2 µg/mL for TGF-β1.

b. We recommend reconstituting ascorbic acid in DMEM/F-12 medium to create a 200 mM stock solution; See "Preparation of 200 mM Ascorbic Acid" at end of this document for important information.

2. Working in a cell culture hood (biological safety cabinet), combine all Microglia Base Medium components in an appropriately sized sterile container. Allow the Cell Culture Medium to equilibrate to room temperature for at least 15 minutes. Do not warm the medium in a 37°C water bath. Culture plates should also be at room temperature prior to use.
3. Prepare pre-diluted Cultrex solution: add 495 µL of **cold** DMEM/F12 into a 55 µL aliquot of frozen Cultrex taken directly from the -80 freezer. Mix to dissolve and store at 4°C until it is time to seed the plate (Step 16).
4. Prepare one 50-mL conical tube: add 3 mL of Microglia Base Medium.
5. Prepare one microcentrifuge tube: add 25 µL of Trypan Blue solution for cell counting.

Day 0 Seeding the Microglia

6. Safety Reminder: the use of safety PPE is recommended during the handling of frozen cryovials. Remove the cryovial from the liquid nitrogen and immediately place in a 37°C water bath. To minimize contamination, avoid submerging the cap. Gently move the vial within the bath to increase the rate of thawing.
7. As soon as the last of the ice melts, which will take ~75-90 seconds, remove the cryovial from the water bath. Disinfect the vial by spraying it with 70% ethanol before transferring it to the cell culture hood.
8. Using a P1000 pipette, transfer 500 µL Microglia Base Medium from the prepared 50-mL conical tube to the cell cryovial at a rate of ~2-3 drops/sec. This process should take about 10 seconds.
9. Gently transfer all contents from the cryovial (~1mL total) back to the same 50-mL conical tube. Add 1 mL of additional Base Medium to the cryovial to collect residual cells and transfer to the same 50-mL tube.
10. Centrifuge cells at 1115 rpm (200xg) for 5 mins.
11. Carefully aspirate supernatant, leaving the cell pellet, and resuspend cells in 950 µL fresh Microglia Base Medium. Mix thoroughly by gently pipetting up and down using a P1000 pipette.
12. Based on the CoA value (*Viable Cells/Vial*), resuspend the cells to a concentration of $1.0x10^6$ live cells/mL by slowly adding additional Base Medium to the existing ~1 mL in the tube.
    a. Example: 2.3 million Viable Cells/Vial is diluted to 2.3 mL total volume.
13. Count the cells: gently swirl the conical tube and pipette up and down 3-5 times to ensure cells are evenly suspended in medium. Transfer 25 µL of cell suspension to the microcentrifuge tube prefilled with 25 µL Trypan Blue solution from Step 5 and pipette up and down a few times to mix. Count the number of viable and dead cells using a hemocytometer. Determine the live cell concentration (live cells/mL) and viability.
14. Calculate volumes needed to make the Microglia Seeding Suspension. A typical seeding density is 20,000 – 30,000 viable cells/100 µL/well for a 96-well plate (~62,500 – 93,750 viable cells/cm^2). Recommended seeding density may vary based on lot number; refer to the CoA for lot-specific seeding information. Dilute the cells to the desired seeding concentration based on the Trypan Blue cell count.

Example of dilution calculations.

Actual Live Cell Concentration	Target Seeding Density	Total Seeding Volume Needed	Volume of Cell Suspension	Volume of Base Medium
Ex: $1.1x10^6$ viable cells/mL *(Step 13)*	20,000 viable cells per 100 *µL/well* = *200,000 cells/mL*	11 mL *(Seeding 1 plate)*	**Mix: 2 mL** *(200,000/mL x 11mL)* / *$1.1x10^6$ cells/mL*	**Add: 9 mL** *(11mL – 2mL)*

15. In a new sterile 50-mL conical tube, mix the calculated volumes of cell suspension and Microglia Base Medium needed to obtain a final volume of 11 mL Microglia Seeding Suspension.
16. Add appropriate volumes of IL-34 (1:1000), TGF-β1(1:1000), M-CSF (1:5000), and *pre-diluted* Cultrex from Step 3 (1:100) to the MG Seeding Suspension per the Seeding Medium recipe.
    a. Example: add 11 µL each of IL-34 and TGF-β1, 2.2 µL of M-CSF, and 110 µL pre-diluted Cultrex solution to 11 mL of MG Seeding Suspension.
17. Mix completely and transfer 100 µL/well (Ex: 20,000 cells/well) of the final Seeding Medium into a PDL-coated 96-well plate using a multi-channel pipette or liquid handler. Do not move or agitate the plate throughout the duration of the seeding process as this may lead to uneven attachment.
18. After seeding, do not immediately transfer the plate to the incubator. let the plate rest for 10 minutes before moving it to allow the cells to settle to the bottom of the well. After 10 minutes, very gently transfer the plate to a to a humidified incubator at 37°C with 5% CO_2. Day of cell plating is designated as Day 0.

    **Note: The entire thawing and plating process should not exceed 1 hour. Post-thaw viability and overall cell health could be severely impacted and lead to an unsuccessful culture if the whole process is too long.*

Day 4 Medium Addition

19. On Day 4 (96 hours after seeding), prepare fresh Maintenance Medium.
20. Gently add 100 µL/well of Maintenance Medium to the entire plate for a total of 200 µL/well.

Day 7 and Onward Medium Changes

21. Change half the medium every 3 days using freshly prepared Maintenance Medium.
22. Gently remove 100 µL/well and slowly add 100 µL/well Maintenance Medium to the entire plate.

The microglia can be maintained viable in culture under the above conditions for at least 3 weeks post-seeding.

Microglia Media Compositions

Step 2		Component	Stock Conc.	Final Conc.	Stock Volume
Microglia Base Medium	1	DMEM/F12 Medium	1X	1X	100 mL
	2	B-27 Supplement	50X	1X	2 mL
	3	N-2 Supplement	100X	1X	1 mL
	4	NEAA	100X	1X	1 mL
	5	Chemically Defined Lipid Concentrate	1X	0.01X	1 mL
	6	GlutaMax	200 mM	1 mM	500 µL
	7	Ascorbic Acid Solution (Step 1b)	200 mM	0.2 mM	110 µL
		Microglia Base Medium stock can be kept for up to 2 weeks at 4°C.			

Step 16		Component	Stock Conc.	Final Conc.	1 Plate Volume	2 Plate Volume	5 Plate Volume
Seeding Medium	1	**Microglia Seeding Suspension** (Step 15)	**1X**	**1X**	**11 mL**	**22 mL**	**55 mL**
	2	IL-34	100 µg/mL	100 ng/mL	11 µL	22 µL	55 µL
	3	TGF-β1	2 µg/mL	2 ng/mL	11 µL	22 µL	55 µL
	4	M-CSF	100 µg/mL	20 ng/mL	2.2 µL	4.4 µL	11 µL
	5	Pre-diluted Cultrex*	1.5 mg/mL	15 µg/mL	110 µL	220 µL	550 µL

* Cultrex must be pre-diluted according to step 3 in the protocol and added to the final seeding suspension right before seeding the plate.

Steps 19, 21		Component	Stock Conc.	Final Conc.	1 Plate Volume	2 Plate Volume	5 Plate Volume
Maintenance Medium	1	**Microglia Base Medium** (Step 2)	**1X**	**1X**	**11 mL**	**22 mL**	**55 mL**
	2	IL-34	100 µg/mL	100 ng/mL	11 µL	22 µL	55 µL
	3	TGF-β1	2 µg/mL	2 ng/mL	11 µL	22 µL	55 µL
	4	M-CSF	100 µg/mL	20 ng/mL	2.2 µL	4.4 µL	11 µL

Preparation of 200 mM Ascorbic Acid Solution

Additional Sterile Materials Required

50-mL Conical Tubes, 0.22 micron Filter, Amber Microcentrifuge Tubes

Critical Information

- Ascorbic Acid must be added *into* the DMEM/F12 (not vice versa) to ensure it dissolves.
- The rate of Ascorbic Acid addition to DMEM/F12 is critical. If added too rapidly, the chemical will fail to go into solution.
- Only mix by inversion. Undissolved Ascorbic Acid will adhere to pipette tips and other implements.
- Ascorbic Acid is pH, temperature, and light sensitive.

Steps

1. Aliquot 17.2 mL of DMEM/F12 into a 50-ml tube.
2. Weigh 1.0 gram of Ascorbic Acid using a separate vessel.
3. **Critical Step**: Gradually transfer Ascorbic Acid ***into*** the tube of DMEM/F12 (no more than 0.2 gram at a time, waiting 10 seconds between each addition).
4. Secure the cap on the tube and invert to mix.
5. Allow the Ascorbic Acid to dissolve into the solution:

a. Place in a 37°C water bath for 30 minutes, inverting the tube every 10 minutes to mix.

b. After 30 minutes check to see if Ascorbic Acid is fully dissolved. If not, invert tube to mix and place capped tube at 4°C overnight.

c. After overnight storage, invert to mix and confirm Ascorbic Acid is completely dissolved.

6. Filter the fully dissolved Ascorbic Acid Solution using a Steriflip filter or equivalent 0.22 µm sterile filter.
7. Aliquot into amber 1.5mL Eppendorf tubes; recommended volumes of 110µl or 550µl.
8. Maintain frozen aliquots at -20°C for up to 6 months. Aliquots are single use. Do not refreeze.